Mini prep (Plasmid DNA purification)

1. Collect 15ml tubes and centrifuge (13,000 rpm, 4°C, 15mins).

2 Resuspend (pipetting) pelleted bacteria cells in 250ul Buffer RES. Move it to new 1.5ml Eppendorf tubes.

3 Add 250ul Buffer LYS and mix gently by inverting the tube, then incubate for 2-3 min at Room temperature (Do not Vortex)

4. Add 350ul Buffer NEU and invert the tube immediately but gently 4-6 times.

5. Centrifuge (13,000 rpm, 4°C, 10mins).

6. Transfer all the supernatant (800ul) to a new 1.5ml Eppendorf tube.

7. Add 600ul of 100% 2-Isopropanol to precipitate the plasmid DNA. Vortex thoroughly.

8. Centrifuge (13,000 rpm, 4°C, 20mins). Carefully discard the supernatant.

9. Add 1ml of 70% ethanol to wash the pellet and centrifuge (13,000 rpm, 4°C, 5mins). Carefully remove ethanol completely from the tube through aspirator.

10. Let the pellet to dry for 5min at room temperature.

11. Dissolve the DNA pellet in an appropriate volume (roughly 25 ul) of TE.